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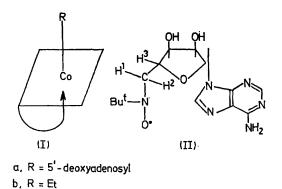
## Light-induced Anaerobic Co<sup>III</sup>-C Homolysis of Aqueous Vitamin B<sub>12</sub> Coenzyme or of Ethylcobalamin; Spin-trapping of the 5'-Deoxyadenosyl or Et Radical

By KEITH N. JOBLIN, ALAN W. JOHNSON, MICHAEL F. LAPPERT,\* and BRIAN K. NICHOLSON (School of Molecular Sciences, University of Sussex, Brighton BN1 9QJ)

Summary Irradiation of vitamin  $B_{12}$  coenzyme or ethylcobalamin (LCo<sup>III</sup>-R) in the presence of a nitroso compound R'NO (R' = Bu<sup>t</sup> or 2,3,5,6-Me<sub>4</sub>C<sub>6</sub>H) under anaerobic conditions in H<sub>2</sub>O (R' = Bu<sup>t</sup>), Me<sub>2</sub>SO, or PhOH-CHCl<sub>3</sub>, affords the spin-trapped nitroxide R'N( $\dot{O}$ )R.

WE report e.s.r. results on the photolysis of vitamin  $B_{12}$  coenzyme (Ia) [and on the related compound ethylcobalamin (Ib)] in an aqueous medium which demonstrate unequivocally that light-induced homolysis of the Co<sup>III</sup>-R bond occurs with formation of the 5'-deoxyadenosyl (or Et) free radical as well as vitamin  $B_{12r}$  (a Co<sup>II</sup> corrinoid). These have implications for the mechanism of  $B_{12}$ -dependent biological isomerisations,<sup>1</sup> and studies using similar techniques are in hand to ascertain whether there is a parallel with the substrate-induced enzymatic reactions.

Irradiation (Pyrex-filtered light from a 250 W highpressure mercury lamp) of a deoxygenated mixture of (Ia)  $(10^{-3} \text{ M})$  and Bu<sup>t</sup>NO  $(10^{-2} \text{M})$  in water at 50° in the cavity of an e.s.r. spectrometer produces the signal of spin-trapped 5'-deoxyadenosyl radical Bu<sup>t</sup>N(O)R (R = 5'-deoxyadenosyl), (II) (see Figure and Table). Ethylcobalamin (Ib)



behaves similarly, giving Bu<sup>t</sup>N(O)Et. The mode of Co-C cleavage appears to be solvent independent; thus irradiation

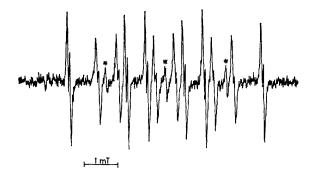


FIGURE. E.s.r. spectrum of  $Bu^{t}N(\dot{O})R$  (R = 5'-deoxyadenosyl) in H<sub>2</sub>O at 50°. Peaks marked\* are due to  $Bu_2^tNO$ .

of (Ia) or (Ib) in either  $PhOH-CHCl_3$  (1:1) or the aprotic solvent Me<sub>2</sub>SO, in the presence of nitrosodurene as spin-trap, similarly afforded the nitroxide ArN(O)R (see Table). In each experiment, the  $B_{12r}$  moiety was characterised by a broad e.s.r. signal at g ca. 2.2, upon freezing the irradiated sample to 173 K.

## TABLE

E.s.r. parameters<sup>a</sup> of the nitroxides  $R'N(\dot{O})R$ 

R′	R	a(N)	$a(\mathrm{H}^{1})$	$a(\mathrm{H}^2)$	$a(\mathrm{H}^3)$
Butb	5'deoxyadenosyl	1.64	1.41	0.81	0.06
Arc	"	1.49	1.49	0.76	е
Ard	"	1.38	1.59	0.56	е
Butb	Et	1.71	$1.13^{1}$		
Are	**	1.51	$1.26^{t}$		

<sup>a</sup> At 50°C, coupling constants in mT. <sup>b</sup> In H<sub>2</sub>O. <sup>c</sup> In CHCl<sub>3</sub>-<sup>d</sup> In Me<sub>2</sub>SO. <sup>e</sup> Not resolved. <sup>f</sup> H<sup>1</sup>=H<sup>2</sup>, giving phenol (1:1). 1:2:1 triplet.

The nitroxides derived from (Ia) were unambiguously identified by the form and line-width variations in the e.s.r. spectrum (see Table). It has been shown that  $\alpha$ -methylene protons (H<sup>1</sup> and H<sup>2</sup>) of a nitroxide with a  $\beta$ -optically active chiral centre R'N( $\dot{O}$ )CH<sub>2</sub>CXYZ (X  $\neq$  Y  $\neq$  Z) are magnetically non-equivalent and thus give rise to a 1:1:1:1 quartet, with selective broadening of the inner pair,<sup>2</sup> rather than the 1:2:1 triplet observed when the  $\beta$ -carbon atom is not chiral. Furthermore, the spectrum of ButN(O)R (R = 5'-deoxyadenosyl) shows a small splitting attributable to hyperfine coupling with the hydrogen (H<sup>3</sup>) on the  $\beta$ -carbon atom.

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<sup>1</sup> Cf. D. Dodd and M. D. Johnson, J. Organometallic Chem., 1973, 52, 1; H. A. O. Hill, in 'Inorganic Biochemistry,' ed. G. I. Eichhorn, Elsevier, Amsterdam, 1973, Vol. 2, 1067; D. J. Cardin, K. N. Joblin, A. W. Johnson, G. Lang, and M. F. Lappert, Biochem. Biophys. Acta, 1974, 371, 44, and references therein. <sup>2</sup> C. Lagercrantz and M. Setaka, Acta Chem. Scand., 1974, B28, 619 and references therein.